

This Page Is Inserted by IFW Operations  
and is not a part of the Official Record

## **BEST AVAILABLE IMAGES**

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

**IMAGES ARE BEST AVAILABLE COPY.**

As rescanning documents *will not* correct images,  
please do not report the images to the  
Image Problem Mailbox.

10



Europäisches Patentamt  
European Patent Office  
Office européen des brevets

11 Publication number:

**0 370 989  
A2**

12

## EUROPEAN PATENT APPLICATION

21 Application number: 89870181.8

51 Int. Cl. **C12N 15/12, C12P 21/02,  
C12N 5/16**

22 Date of filing: 20.11.89

30 Priority: 21.11.88 US 274061  
10.07.89 US 377037

43 Date of publication of application:  
30.05.90 Bulletin 90/22

64 Designated Contracting States:  
AT BE CH DE ES FR GB GR IT LI LU NL SE

71 Applicant: MONSANTO COMPANY  
800 North Lindbergh Boulevard  
St. Louis Missouri 63167(US)

72 Inventor: Keck, Pamela Jean  
1420 Willowbrook Cove  
St. Louis Missouri 63146(US)  
Inventor: Feder, Joseph  
935 Mulberry Lane  
University City Missouri 63130(US)  
Inventor: Connolly, Daniel Thomas  
6 Chapel Court  
Manchester Missouri 63021(US)  
Inventor: Olander, Jitka Vera  
7114 Kingsbury Boulevard  
University City Missouri 63130(US)

74 Representative: Lunt, John Cooper et al  
Monsanto Services International S.A. Patent  
Department Avenue de Tervuren 270/272  
LBox 21  
B-1150 Brussels(BE)

84 Human vascular permeability factor.

87 A cDNA having a base sequence for human vascular permeability factor has been cloned and characterized and the amino acid sequence of the human vascular permeability factor protein has been determined.

**EP 0 370 989 A2**

## HUMAN VASCULAR PERMEABILITY FACTOR

Background of the Invention

This invention relates to a novel human vascular permeability factor and to a cDNA clone representing the full size human vascular permeability factor protein.

Vascular permeability factors (VPFs) are proteins originally obtained from a variety of tumors which cause a rapid and reversible increase in blood vessel permeability when nanogram amounts are injected under the skin of a warm blooded mammal. VPF activity has been found in tumor ascites fluid from guinea pigs, hamsters and mice and is secreted by these tumors and a variety of tumor cell lines *in vitro* according to Senger et al., *Science* 219, 983-985 (1983).

In U.S. Patent 4,456,550, a purified VPF is described which has the following characteristics:

(a) in an aqueous solution (0.01 M  $\text{Na}_2\text{PO}_4$ , pH 7) whose concentration of NaCl is varied linearly, VPF is eluted from a heparin-Sepharose chromatography column in a peak centered at 0.4 NaCl;

(b) in an aqueous solution of  $\text{Na}_2\text{PO}_4$ , pH 7.0, whose concentration is varied linearly, VPF is eluted from a hydroxylapatite column in a peak centered at 0.25 M  $\text{Na}_2\text{PO}_4$ ; and

(c) when subjected to SDS gel electrophoresis in a polyacrylamide slab gel (0.375 M tris-HCl, pH 8.8, 0.1% SDS) at 35 milliamps and 4° C., VPF is localized to a region corresponding to a molecular weight between 34,000 and 45,000 daltons.

VPF of the foregoing characteristics was thus purified about 1800 fold from serum-free conditioned medium of guinea pig tumor cell culture or 10,000 fold from ascites fluid by a series of steps consisting of:

(a) affinity chromatography with a column of heparin-Sepharose;

(b) chromatography with a column of hydroxylapatite; and

(c) sodium dodecylsulfate/polyacrylamide gel electrophoresis. According to said patent, as little as 200 ng ( $5 \times 10^{-12}$  moles) of this purified VPF increased the vascular permeability equivalent to 1.25  $\mu\text{g}$  ( $4 \times 10^{-9}$  moles) of histamine. Histamine is a standard permeability mediator described by Miles and Miles, *J. Physiol.* 118, 228-257 (1952). The VPF is said to have therapeutic value insofar as it enables blood nutrients to reach tissue with increased need for nutrients, as in wound healing.

According to Folkman and Klagsbrun, *Science* 235, 442-447 (1987), VPF causes leakage of proteins, including fibrinogen, from blood vessels, thereby initiating the formation of a fibrin gel which, in turn, may play a role in angiogenesis. See also Dvorak et al., *J. Immunol.* 122(1), 166-174 (1979); Dvorak, *N. Engl. J. Med.* 315, 1650-1659 (1986); Kadish et al., *Tissue & Cell* 11, 89 (1979); and Dvorak et al., *J. Natl. Cancer Inst.* 62, 1459-1472 (1979).

In an application assigned to a common assignee, a method of stimulating endothelial cell growth is provided which comprises subjecting said cells to a growth stimulating amount of a highly purified VPF. The highly purified VPF derived from guinea pig tumor cells has the following characteristics:

(a) it has a M<sub>r</sub> about 34,000 - 40,000 as determined by sodium dodecylsulfate polyacrylamide gel electrophoresis (SDS/PAGE);

(b) it is a disulfide-linked protein dimer;

(c) it has a N-terminal amino acid sequence as follows:

1 AlaProMetAlaGluGlyGluGlnLysProArgGluValValLys

16 PheMetAspValTyrLysArgSerTyrCysArgProIleGluMet

31 LeuValAspIlePheGln; and

(d) it exhibits substantial mitogenic activity to endothelial cells in culture.

The foregoing highly purified guinea pig VPF, also referred to as gVPF, was isolated from serum-free conditioned culture medium of guinea pig tumor cells in a series of steps comprising:

(a) affinity chromatography of said conditioned culture medium with a column of heparin-Sepharose

CL-6B;

(b) cation exchange chromatography of the VPF active fractions from said affinity chromatography with a TSK SP-5-PW column;

(c) high performance liquid chromatography (HPLC) of the VPF active fractions from said cation exchange chromatography with a Vydac C<sub>4</sub> reversed phase HPLC column; and

(d) HPLC of the VPF active fractions from said C<sub>4</sub> HPLC with a Vydac C<sub>18</sub> reversed phase HPLC column.

In an application assigned to a common assignee, a method of producing antibodies against gVPF is provided in which certain peptide fragments of gVPF are used as immunogens.

Lobb et al., *Int. J. Cancer* 38, 473-478 (1985), describe a partially purified VPF from a human adenocarcinoma cell line HT-29 having a molecular weight of 45,000. This VPF, however, does not bind to immobilized heparin as does the VPF derived from guinea pig tumor cell material by Senger and Dvorak.

Senger et al., *Cancer Res.* 48, 5629-5632 (1988), describe the production of VPF from a variety of human tumor cell lines, namely human osteogenic sarcoma, bladder sarcoma, cervical carcinoma and fibrosarcoma cell lines. However, none of these human cell lines were found to be as active as the guinea pig cell line 10 for the production of VPF.

#### Brief Description of the Invention

20

In accordance with the present invention, a cDNA clone representing the full size human vascular permeability factor (hVPF) has been developed. This clone, VPF-4, contains about 3550 base pairs (bp) with an apparent 5'-noncoding region of about 1300 bp, an open reading frame of 845 bp, a stop codon and a 3'-noncoding region of about 1600 bp.

The cDNA sequence encodes a hVPF protein of 215 amino acids. This protein sequence includes a putative signal peptide of 26 amino acids beginning with a methionine residue positioned 26 amino acids upstream to the NH<sub>2</sub>-terminus, followed by the 189 amino acid mature protein.

The hVPF contains a potential N-glycosylation site at Asn 75 and 16 cysteine residues. A zinc-finger like region is noted at amino acids 57-60.

Within the hVPF sequence, four amino acid segments correspond to the previously sequenced segments from the U-837 secreted protein described in United States patent application Ser No. 07/274,061 which is one of the applications from which the present application claims priority. These include the first 10 amino acids of the NH<sub>2</sub>-terminus, namely

35

<sup>1</sup> A P M A E G G G Q <sup>10</sup> N,

40 and three internal peptide fragments which correspond to amino acid residues:

(a) <sup>17</sup> F M D V Y Q <sup>23</sup> R,

45

(b) <sup>33</sup> V D I F Q E Y P D E I E Y, and

50

(c) <sup>46</sup> I F K P S C V P L M R. <sup>56</sup>

The cDNA sequence and corresponding predicted amino acid sequence of the hVPF is shown in the following 1195 bp sequence which includes portions of the 5'- and 3'- noncoding regions. Nucleotides are numbered in the lefthand column, beginning with No. 1. Amino acids are numbered from above. The predicted amino acid sequences from the cDNA clone for amino acid positions 1-18, 17-23, 33-45 and 46-56 exactly match amino acid sequences determined by amino acid sequencing of the N-terminus and of three different tryptic peptides. These sequences are underlined. The single N-glycosylation site at Asn-75

is boxed. The stop codon is designated with three asterisks.

5

10

15

20

25

30

35

40

45

50

55

1 GCGCAGACAG TGCTCCAGCG CGCGCGCTCC CCAGCCCTGC CCGGCCTCGG  
 5 51 GCCGGGAGGA AGAGTAGCTC GCCGAGGCGC CGAGGAGAGC GGGCCGCCCC  
 10 101 ACAGCCCGAG CCGGAGAGGG ACGCGAGCCG CCGCCCCCGG TCGGGCCTCC  
 15 151 -26 -20  
 Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu  
 GAAACC ATG AAC TTT CTG CTG TCT TGG GTG CAT TGG AGC CTT  
 18 193 -10  
 Ala Leu Leu Leu Tyr Leu His His Ala Lys Trp Ser Gln Ala  
 GCC TTG CTG CTC TAC CTC CAC CAT GCC AAG TGG TCC CAG GCT  
 20 235 +1 10  
 Ala Pro Met Ala Glu Gly Gly Gly Gln Asn His His Glu Val  
 GCA CCC ATG GCA GAA GGA GGA GGG CAG AAT CAT CAC GAA GTG  
 25 277 20  
 Val Lys Phe Met Asp Val Tyr Gln Arg Ser Tyr Cys His Pro  
 GTG AAG TTC ATG GAT GTC TAT CAG CGC AGC TAC TGC CAT CCA  
 30 319 30 40  
 Ile Glu Thr Leu Val Asp Ile Phe Gln Glu Tyr Pro Asp Glu  
 ATC GAG ACC CTG GTG GAC ATC TTC CAG GAG TAC CCT GAT GAG  
 35 361 50  
 Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu Met Arg  
 ATC GAG TAC ATC TTC AAG CCA TCC TGT GTG CCC CTG ATG CGA  
 40 403 60 70  
 Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro  
 TGC GGG GGC TGC TGC AAT GAC GAG GGC CTG GAG TGT GTG CCC  
 45 445 80  
 Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys  
 ACT GAG GAG TCC AAC ATC ACC ATG CAG ATT ATG CGG ATC AAA  
 50 487 90  
 Pro His Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln  
 CCT CAC CAA GGC CAG CAC ATA GGA GAG ATG AGC TTC CTA CAG  
 55 529 100 110  
 His Asn Lys Cys Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg  
 CAC AAC AAA TGT GAA TGC AGA CCA AAG AAA GAT AGA GCA AGA  
 60 571 120  
 Gln Glu Lys Lys Ser Val Arg Gly Lys Gly Lys Gly Gln Lys  
 CAA GAA AAA AAA TCA GTT CGA GGA AAG GGA AAG GGG CAA AAA  
 65 613 130 140  
 Arg Lys Arg Lys Lys Ser Arg Tyr Lys Ser Trp Ser Val Pro  
 CGA AAG CGC AAG AAA TCC CGG TAT AAG TCC TGG AGC GTT CCC

150  
 655 Cys Gly Pro Cys Ser Glu Arg Arg Lys His Leu Phe Val Gln  
 TGT GGG CCT TGC TCA GAG CGG AGA AAG CAT TTG TTT GTA CAA  
 160  
 697 Asp Pro Gln Thr Cys Lys Cys Ser Cys Lys Asn Thr Asp Ser  
 GAT CCG CAG ACG TGT AAA TGT TCC TGC AAA AAC ACA GAC TCG  
 170 180  
 739 Arg Cys Lys Ala Arg Gln Leu Glu Leu Asn Glu Arg Thr Cys  
 CGT TGC AAG GCG AGG CAG CTT GAG TTA AAC GAA CGT ACT TGC  
 15  
 781 Arg Cys Asp Lys Pro Arg Arg \*\*\*  
 AGA TGT GAC AAG CCG AGG CGG TGA GCCGGGCAGG AGGAAGGAGC  
 825 CTCCCTCAGG GTTTCGGGAA CCAGATCTCT CACCAGGAAA GACTGATACA  
 875 GAACGATCGA TACAGAAACC ACGCTGCCGC CACCACACCA TCACCATCGA  
 925 CAGAACAGTC CTTAATCCAG AAACCTGAAA TGAAGGAAGA GGAGACTCTG  
 975 CGCAGAGCAC TTTGGGTCCG GAGGGCGAGA CTCCGGCGGA AGCATTCCCCG  
 1025 GGCGGGTGAC CCAGCACGGT CCCTCTTGGG ATTGGATTCTG CCATTTTATT  
 1075 TTTCTTGCTG CTAAATCACC GAGCCCGGAA GATTAGAGAG TTTTATTCT  
 1125 GGGATTCTCTG TAGACACACC CACCCACATA CATACATTTA TATATATATA  
 1175 TATTATATAT ATATAAATTA A

The gene coding for the hVPF of this invention can be cloned into and expressed in prokaryotic and eukaryotic hosts. For example, active hVPF protein can be expressed in a prokaryotic host such as *E. coli* and a eukaryotic host such as C-127 mouse cells or Chinese hamster ovary (CHO) cells by operably inserting the hVPF coding sequence in replicable expression vectors or plasmids. For example, it can be inserted into a suitable plasmid such as pML for production in *E. coli* and the bovine papilloma virus (BPV) vector for production in mouse cells or a shuttle vector which can replicate in both prokaryotic and eukaryotic cells. In a preferred embodiment, the gene coding for the hVPF was cloned into and expressed from C-127 mouse cells. The excreted protein was determined by the assay of Miles and Miles, *supra*, (hereinafter also referred to as the Miles assay).

The original source material for the production of hVPF is the human histiocytic lymphoma cell line U-937. This cell line was originally established from cells from the pleural effusion of a patient with diffuse histiocytic lymphoma as reported by Sundstrom and Nilsson, *Int. J. Cancer* 17, 565-577 (1976). These cells are widely distributed as evidenced by publications and are also readily available to the public in an unrestricted culture deposit from the American Type Culture Collection, Rockville, Maryland, under accession number ATCC CRL 1593. Further background on these cells can be had by reference to *J. Exp. Med.* 143, 1528-1533 (1976); *Nature* 279, 328-331 (1979); and *J. Immunol.* 125, 483-485 (1980).

A recent report on the use of U-937 cells to produce VPF-like activity was made by Beck and Habicht, *J. Leukocyte Biol.* 42, 588 Absts., Dec. 1987. However, the activity was not purified and chemical characterization or identity was not disclosed.

The human VPF produced by U-937 cells was originally identified by its inhibition and binding by rabbit polyclonal antibodies to guinea pig VPF. A rabbit polyclonal antiserum to guinea pig VPF inhibited the permeability activity produced by U-937 cells as determined by the of Miles assay. This U-937 generated VPF activity was about 70% to 80% removed by binding to immunoadsorbents produced with protein A-

Sepharose which had been reacted with the rabbit polyclonal antiserum to guinea pig VPF.

A suitable process for the production of human VPF comprises growing cells derived from the human histiocytic lymphoma cell line U-937 in serum-free nutrient culture medium at about 35° to 38° C for a sufficient time to elaborate VPF and isolating the resulting VPF from the spent cells or the cell culture conditioned medium.

A preferred method of isolating the human VPF from the cell culture conditioned medium of the U-937 cells described in said co-pending application comprises the following steps:

(a) cation exchange chromatography of said conditioned cell culture medium, for example with a column of CM-cellulose, CM-Sephadex®, Amberlite® IR-120H or S-Sepharose Fast Flow cation exchanger;

(b) metal affinity chromatography of the VPF active fractions from said cation exchange chromatography, for example with a  $\text{Cu}^{2+}$ ,  $\text{Zn}^{2+}$  or  $\text{Ni}^{2+}$ /iminodiacetic acid (IDA)/Sepharose column; and

(c) reverse phase HPLC of the active VPF fractions from said method affinity chromatography, for example with a  $\text{C}_4$  or  $\text{C}_{18}$  reverse phase HPLC column.

The thus purified human VPF was found to be a protein of M<sub>r</sub> 34,000-42,000. When subjected to N-terminal amino-acid sequence analysis, it was found to have a distinct and novel structure whereby it differed from gVPF in four of the first ten amino acid positions.

The purified hVPF was active in promoting vessel leakage at a dose of 22 ng ( $5.5 \times 10^{-13}$  Moles) upon intradermal injection into guinea pigs. This highly purified hVPF thus is 9 times more potent than the gVPF described in U.S. Patent 4,456,550. Another advantage of hVPF of this invention is its human origin which is indicative of potential use as a human therapeutic compared to other agents of lesser purity or derived from guinea pig or other animals such as would cause immunological reactions.

#### Detailed Description of the Invention

While the specification concludes with claims particularly pointing out and distinctly claiming the subject matter regarded as forming the present invention, it is believed that the invention will be better understood from the following detailed description of preferred embodiments of the invention taken in conjunction with the accompanying drawings in which:

FIG. 1 is a schematic representation which shows the stepwise isolation of the human VPF (hVPF) from the conditioned cell culture medium of U-937 cells in one embodiment of the invention.

FIG. 2 is a graphical representation which shows the elution patterns in the stepwise purification of the hVPF in four panels A, B, C and D in the embodiment of FIG. 1 as follows:

A.) Cation Exchange Chromatography of hVPF. Serum-free conditioned medium from U-937 cells (8 L of 8-fold concentrate) was adjusted to pH 7.0 and loaded at a flow rate of 60 ml/hr onto an S-Sepharose column (5 x 45 cm) equilibrated in 0.01 M sodium phosphate, pH 7.0. A linear gradient from 0.2 M to 0.8 M NaCl in the same buffer was used to elute hVPF.

B.) Metal Affinity Chromatography of hVPF. The active eluate from the cation exchange column was concentrated to 20 ml by ultrafiltration and loaded onto a Sepharose (Fast Flow)/IDA/ $\text{Cu}^{2+}$  column equilibrated in 0.01 M sodium phosphate, pH 7.0, 2 M sodium chloride, 0.5 M imidazole. hVPF was eluted with a linear gradient of imidazole as shown.

C.) RP-HPLC of hVPF. The active eluate from the metal affinity column was loaded onto a  $\text{C}_{18}$  RP-HPLC column equilibrated with 0.05% trifluoroacetic acid (TFA) in water and eluted at 1 ml/min with a gradient of acetonitrile as shown.

D.) Sodium Dodecylsulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) of RP-HPLC Fractions. Aliquots were removed from fractions around the activity peak and analyzed by electrophoresis without reducing agent. Standards were first reduced with  $\beta$ -mercaptoethanol.

FIG. 3 shows the ELISA of hVPF and gVPF. Rabbit anti-gVPF IgG was used to coat the polystyrene wells of microtiter plates. Various amounts of either hVPF (lower panel) or gVPF (upper panel) were allowed to bind overnight, and the amount of bound antigen detected with biotin-anti-gVPF IgG followed by HRP-avidin. Wells were developed with HRP substrate, the absorbance was read at 490 nm. Wells containing readings that were off scale were diluted and re-read in the linear range of the Bio-Tek microplate reader; the absorbances given are corrected for dilution. Different x-axis scales were used for hVPF and gVPF. Early and late bleed refers to both primary and secondary biotin-conjugated IgGs prepared from sera collected at the fourth bleed (after 3 immunizations) and at the eleventh bleed (after 5 immunizations), respectively.

FIG. 4 shows the structure of plasmid hVPF/pUC9 of about 6200 bp. This plasmid, also designated



pMON3044, contains the hVPF cDNA. The heavy arrow represents the location of the coding region for hVPF within the 3.5 kb cDNA.

Plasmid pUC9 is a commercially available versatile vector that contains a multiple cloning site and a gene that confers ampicillin (AMP) resistance. The plasmid produces the  $\alpha$ -peptide of  $\beta$ -galactosidase which complements the lac deletion mutation in *E. coli* strains such as DH5a, JM83 and TB1.

Standard biochemical nomenclature is used herein in which the nucleotide bases of DNA or RNA are designated as adenine (A); thymine (T); guanine (G); and cytosine (C). N denotes any of these nucleotides. As is conventional for convenience in the structural representation of a DNA nucleotide sequence, only one strand is usually shown in which A on one strand denotes T on its complement and G denotes C. All sequences are written 5' to 3'. Amino acids are shown either by three letter or one letter abbreviations as follows:

Abbreviated Designation		Amino Acid
A	Ala	Alanine
C	Cys	Cysteine
D	Asp	Aspartic acid
E	Glu	Glutamic acid
F	Phe	Phenylalanine
G	Gly	Glycine
H	His	Histidine
I	Ile	Isoleucine
K	Lys	Lysine
L	Leu	Leucine
M	Met	Methionine
N	Asn	Asparagine
P	Pro	Proline
Q	Gln	Glutamine
R	Arg	Arginine
S	Ser	Serine
T	Thr	Threonine
V	Val	Valine
W	Trp	Tryptophan
Y	Tyr	Tyrosine

Commonly available restriction endonucleases shown in Example 4 and FIG. 4 have the following restriction sequences and (indicated by arrows) cleavage patterns.

5	BamH1	GGATCC CCTAGG	Xma1	CCCGGG GGGCCC
10	EcoR1	GAATTC CTTAAG		
15	Hind3	AAGCTT TTCGAA		
20	Pvu1	CGATCG GCTAGC		
25	Sa11	GTCGAC CAGCTG		

The U-937 cells can be cultured in well-known cell culture media such as basal medium Eagle's (BME), Dulbecco's modified Eagle medium (DMEM), medium 199, RPMI 1640 medium, and the like cell culture media such as described in detail by H. J. Morton, *In Vitro* 6, 89-108 (1970). These conventional culture media contain known amino acids, mineral salts, vitamins, hormones and carbohydrates. They are also frequently fortified with mammalian sera such as fetal bovine serum (FBS). Other components which can be used in the media are bovine serum albumin (BSA), growth factors such as transferrin and insulin, protein hydrolysates such as lactalbumin hydrolysate, tryptone, tryptose and peptone, as well as lipids, surfactants and the like materials. The U-937 cells preferably are cultured in serum-free media for the production of hVPE.

Methods for the large scale growth of mammalian cells are well-known and these methods can be used for the culture of the U-937 cells defined herein. Such methods are described, for example, by Tolbert et al., Biotech. Bioeng. XXIV, 1671-1679 (1982); Tolbert and Feder, Ann. Rept. Ferm. Proc. Vol. 6, Ch. 3, pp. 35-74 (1983); Harakas, ibid. Vol. 7, Ch. 7, pp. 159-211 (1984); and references cited in said publications. U.S. Pat. Nos. 4,166,768; 4,289,854; and 4,537,880 disclose particularly useful methods and apparatus for the large scale growth and maintenance of cells for the production of proteinaceous materials. The disclosures in said patents are incorporated herein by reference. The methods and apparatus disclosed therein can be used for the culture of the U-937 cells defined herein.

The cells also can be cultured on a large scale basis in nutrient medium at 37° C. in agitated suspension culture as described in U.S. Pat. No. 4,289,854 and, after a suitable growth period, can be maintained in the static maintenance reactor described in U.S. Pat. No. 4,537,860 in which the medium is supplemented with 0.5% lactalbumin hydrolysate.

Although purification of the hVPF from the spent culture media can employ various known procedures for the separation of proteins such as, for example, salt and solvent fractionation, adsorption with colloidal materials, gel filtration, ion exchange chromatography, affinity chromatography, immuno affinity chromatography, electrophoresis and high performance liquid chromatography (HPLC), the above described three-step chromatographic method is preferred. Suitable metal affinity chromatography procedures are illustrated by Sulkowski, Trends Biotech. 3, 1-7 (1985).

In a preferred process the U-937 cells are subcloned or passaged through nude mice to improve the yield of hVPF. Such treatment of the cells has provided for the production of hVPF in quantities of up to 200 to 800 ng/ml of the conditioned medium. This is equivalent to or greater than the gVPF levels reported for the guinea pig cell Line 10.

The U-937 cells have been passaged through nude mice by injection of  $1 \times 10^7$  cells into the mouse

peritoneum. Nude mice are an immunodeficient species in which human cells can grow and not be rejected. After about 3 to 4 weeks, the mice were sacrificed and soft tumors were removed from their abdomens. The tumors were mechanically dissociated into cells and these U-837 cells were again put into culture.

The following examples will further illustrate the invention although it will be appreciated that the invention is not limited to these specific examples or the details described therein.

### Example 1

## MATERIALS AND METHODS

### Growth of U-837 Cells.

U-837 cells originally obtained from the American Type Culture Collection (ATCC) were subcloned in soft agar and selected for fast growth. One of these clones was selected for scale-up, but other clones, and even uncloned ATCC cells, also produced hVPF. The serum-free medium used contained the following components: RPMI 1640, DME (high glucose), Ham's F12 in a 1:1:1 ratio; HEPES (25 mM, pH 7.10-7.15); glutathione (1 mM); ethanolamine (20  $\mu$ M); selenium (30 nM);  $\text{NaHCO}_3$  (2 mM);  $\text{CuSO}_4$  (5 nM);  $\text{NH}_4\text{VO}_3$  (5 nM);  $\text{ZnSO}_4$  (0.5  $\mu$ M);  $\text{MnSO}_4$  (0.5 nM);  $\text{FeSO}_4$  (4  $\mu$ M); bovine serum albumin, Miles "Pentex" (100  $\mu$ g/ml); iron rich transferrin, Miles (5  $\mu$ g/ml); bovine insulin (10  $\mu$ g/ml); ExCyte, Miles-lipid fraction (0.1% v/v); F-88 Pluracol® (0.05% w/v). The volume was adjusted to yield an osmolarity of 280 mOsm. The doubling time was about 50-60 hours in this medium, whereas it was only 35-40 hours in medium containing 2% fetal bovine serum (FBS).

Cells were scaled-up in the serum-free medium from T-flasks into roller bottles and then into small spinners. A 12 L spinner was then used to inoculate a 14 L perfusion chemostat which was perfused at a rate of approximately 3 ml medium/hour/ml of total packed cells. The culture was subsequently transferred to a 100 L perfusion chemostat, which was perfused under limiting nutrient conditions (1.5-2.0 ml medium/hour/ml packed cells, or 0.1-0.15 ml/day/million cells). Cells were recycled to the reactor using an AG Technology hollow fiber-cartridge (model CFP-4-E-6, 0.4 micron). Cell density ranged from  $1.0 \times 10^6$  to  $4.8 \times 10^6$  viable cells/ml, 3.0 to 23 ml/L of packed cells, and viability ranged from 64% to 84%. The production run in the 100 L reactor lasted 24 days, during which time a total of 1000 L of serum free conditioned medium was produced.

The permeate from the perfusion reactor was collected and stored at 4° C. Concentration was performed in 200-300 L lots on an Amicon DC-30 ultrafiltration apparatus with three low protein binding 10 kDa cutoff spiral cartridges (Amicon S10Y10) operated in parallel. A concentration of about 6 fold was achieved, including a phosphate buffered saline (PBS) wash of the cartridges and equipment that was pooled with the concentrate. The concentrate was stored at -20° C.

### Vascular Permeability Assay.

A Miles-type permeability assay (Miles and Miles, *supra*) was used to detect hVPF. Hairless guinea pigs (IAF/HA-HO, Charles River, Wilmington MA) were anesthetized by inhalation of methoxyflurane (Metofane®, Pitman-Moore, Inc.). A 1 ml volume of 0.5% (w/v) Evan's blue dye (Sigma Chemical Co.) prepared in sterile saline for injection (Abbott Laboratories) was injected intracardially into the circulation. Samples for hVPF determination were prepared at appropriate dilutions in saline or phosphate buffered saline, and 200  $\mu$ l volumes injected intradermally into sites on the back of the guinea pig. The presence of hVPF was indicated by an intense blue spot at the site of the injection where dye (bound to serum protein) had leaked from the circulation into the tissues.

### Cation Exchange Chromatography.

Six liters of six-fold concentrated conditioned medium was adjusted to pH 7.0 with acetic acid and passed through a column (5 cm X 44 cm) of S Sepharose Fast Flow (Pharmacia) cation exchange gel equilibrated with 0.01 M sodium phosphate, pH 7.0. At 4° C a significant portion of the permeability enhancing activity passed through the column, but at ambient temperature (25° C) 50 to 70% of the activity became bound to the column. This step was therefore regularly performed at room temperature. Sodium azide (0.01% w/v) was added to all buffers. Flow rates for loading and elution were 10 ml per minute. After loading, the column was washed with 900 ml of 0.01 M sodium phosphate, pH 7.0, and then eluted with a 2.3 L linear gradient containing from 0.2 M to 0.8 M sodium chloride in the same buffer. Between runs, the column was washed with 0.1 M sodium hydroxide before re-equilibrating with 0.01 M sodium phosphate, pH 7.0.

#### Metal Affinity Chromatography.

Metal affinity chromatography was performed using a copper-iminodiacetic acid complex covalently linked to agarose via a spacer arm. The gel was synthesized in two steps by first adding an epoxide-containing spacer arm to the agarose, and then reacting the activated gel with iminodiacetic acid.

Highly cross-linked agarose (Sepharose Fast Flow, Pharmacia) was repeatedly washed with distilled water to remove all buffers and preservatives and then dried by suction. About 100 g (100 ml) of this damp gel was suspended in 60 ml distilled water, and then 40 ml freshly prepared 2.5 M NaOH solution was added to the gently stirring agarose suspension. Then 100 ml diethyleneglycol diglycidyl ether, prepared as described by Gu et al., *Synthesis*, 849-851 (1983), was added, and the mixture was gently stirred at 30° C for 16 hours. The activated gel was repeatedly washed with distilled water to remove the excess epoxide and base. The washed, suction-dried gel contained 70 micromoles active epoxide groups per mL gel. The activated gel was stored in distilled water at 4° C and generally used within 24 hours of preparation.

About 100 ml (100 g) of the activated Sepharose Fast Flow was washed with distilled water, dried by suction, and suspended in 100 ml of 1.0 M  $\text{Na}_2\text{NH}(\text{CH}_2\text{CO}_2)\cdot\text{H}_2\text{O}$  solution, which was adjusted to pH = 11.0. This mixture was gently stirred at 65° C for 24 hours and then repeatedly washed with distilled water to remove excess ligand. The functionalized gel was stored in ethanol/water (25/75 v/v) at 4° C until ready for use. Titration with thiosulfate showed the absence of epoxide groups, so capping with ethanolamine was deemed unnecessary. To determine the metal binding capacity of the gel, 10 ml of suction-dried gel was saturated with excess 50 mM  $\text{Cu}(\text{ClO}_4)_2$  and then carefully washed with distilled water. Finally, the bound copper was removed with an excess of 50 mM  $\text{Na}_2\text{H}_2\text{EDTA}$ . Using standardized copper-EDTA solutions for comparison, the copper content was photometrically determined to be 43 micromoles Cu per milliliter of damp, suction-dried gel.

Chromatography was performed in a glass column (1.0 x 13 cm.) containing 10 ml of gel. The gel was charged with a 50 mM solution of  $\text{Cu}(\text{ClO}_4)_2$ , pH = 4.5, and then saturated with a buffered imidazole solution (20 mM imidazole + 2 M NaCl + 50 mM  $\text{NaH}_2\text{PO}_4$ , pH 7.0). After thoroughly washing with 1.0 M NaCl, the column was then equilibrated with the starting buffer (50 mM  $\text{NaH}_2\text{PO}_4$ , pH 7.0, 2 M NaCl, 0.5 mM imidazole). The fractions from the cation exchange step containing permeability enhancing activity were pooled and concentrated from a volume of about 700 ml to 20 ml using an Amicon YM5 membrane. Upon concentration, a protein precipitate usually formed that could be removed by filtration. The sample was applied to the column at ambient temperature (25° C) and eluted at a flow rate of 0.5 ml/min over 500 minutes with a linear gradient of imidazole (0.5 mM to 60 mM) in 50 mM  $\text{NaH}_2\text{PO}_4$ , pH 7.0, 2 M NaCl.

#### Reverse-Phase HPLC.

Reverse-phase HPLC (RP-HPLC) was performed using a 4.6 mm x 25 cm Vydac column (Separations Group) containing 5  $\mu\text{M}$  packing with 330 angstrom pore size. The mobile phases were: "A", 0.05% trifluoroacetic acid (TFA) in water and "B", 0.05% TFA in acetonitrile. After loading the sample, the column was washed with "A" until the absorbance again reached baseline, and then eluted with the following linear gradients: 0% to 20% "B" over 20 minutes, 20% to 40% "B" over the next 80 minutes, and then 40% to 100% "B" over the next 20 minutes. All flow rates were 1 ml/min. Fractions were collected in siliconized glass tubes.

#### N-Terminal Amino Acid Sequence Analysis.

Automated Edman degradation chemistry was used to determine N-terminal amino acid sequence. An Applied Biosystems, Inc., model 470A gas phase sequencer (Foster City, CA) was employed for the degradations [Hunkapiller, et al, *Methods Enzymol.* **91**, 399-413 (1983)]. The respective PTH-amino acid derivatives were identified by RP-HPLC analysis in an on-line fashion employing an Applied Biosystems, Inc., Model 120A PTH Analyzer fitted with a Brownlee 2.1 mm I.D. PTH-C<sub>18</sub> column. Yields of PTH amino acids were determined by comparison with an external standard mixture. The average repetitive yield was calculated by linear regression analysis of the log pmolar yield versus cycle number plot.

Similarities between the obtained sequence and known sequences were investigated using the computer program FAST A [Pearson and Lipman, *Proc. Natl. Acad. Sci. USA* **85**, 2444-2448 (1988)]. Similarity searches were also performed against the National Biomedical Research Foundation (NBRF) protein sequence data base [Sidney et al., *Nucleic Acids Res.* **16**, 1869-1871 (1988), Release 17, June 1988] and the translated GENBANK DNA base [Bilofsky and Burks, *Nucleic Acids Res.* **16**, 1861-1864 (1988), Release 56, June 1988].

Tryptic peptides were prepared from reduced and alkylated hVPF. hVPF (1 nmole) was dissolved in 100 µl of 0.5 M Tris HCl, pH 8.5, 8 M guanidine-HCl, 1 mM EDTA, and 5 mM dithiothreitol. The solution was incubated for 30 minutes at 37°C before the addition of sodium iodoacetate (to a final concentration of 5 mM) and incubation at 4°C overnight. After dialyzing against 2 M guanidine HCl, 0.01 M Tris-HCl, pH 8.5, and then 0.1 M ammonium bicarbonate, 1 µg of TPCK treated trypsin (Sigma Chemical Co., St. Louis, MO) was added and the solution incubated overnight at 37°C. Peptides were separated by RP-HPLC using a Nucleosil C<sub>18</sub>, 5 micron, 100 Å, 4.6 x 250 mm column (Macherey-Nagel, Inc.). The flow rate was 1 ml/min at room temperature. A linear gradient of 0 to 90% acetonitrile in 0.1% trifluoroacetic acid run over 270 minutes was used for elution.

#### Electrophoresis.

Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) was carried out on 10-15% gradient polyacrylamide gels using a Pharmacia PhastSystem. The buffer systems and the silver staining protocol were those that were recommended by the manufacturer.

#### Antibodies and Immunoassays.

Rabbit polyclonal antiserum to gVPF (designated F001) was prepared by immunizing a New Zealand White (NZW) rabbit with repeated injections of gVPF. The first injection was in Complete Freund's Adjuvant followed by boosts in Incomplete Freund's Adjuvant. gVPF was purified by preparative SDS-PAGE [Senger et al, *Science* **219** 983-985 (1983)] following purification by the method of Senger et al. *Fed. Proc.* **46**, 2102 (1987).

Sandwich enzyme linked immunosorbent assays (ELISA) for human and guinea pig VPF were performed as described below. The IgG fraction of F001 antiserum was purified using adsorption onto protein A-Sepharose. 500 ng/well of the IgG obtained from an 11th bleed serum was coated onto polystyrene microtiter plates for 3 hours. Then various known concentrations of human and guinea pig VPF were allowed to bind overnight. The concentrations were estimated from the maximal dilution which still produced a detectable response in the Miles permeability assay. This concentration was 50 ng/ml of VPF. The amount of bound antigen was detected with 500 ng/well biotin-a-gVPF IgG (11th bleed F001 antiserum) for 2 hours followed by 1/2000 dilution of horseradish peroxidase (HRP)-avidin (Cappel Labs) for 90 minutes. Wells were developed with the HRP substrate o-phenylenediamine-2HCl plus H<sub>2</sub>O<sub>2</sub> and the absorbance was read at 490 nm in a BioTek reader.

It had been observed that the earlier bleeds of rabbit F001 contained a higher concentration and/or higher affinity of antibodies cross-reactive with hVPF compared to later bleeds. Therefore, an ELISA for hVPF was also performed with an early bleed (4th bleed) antiserum. It was performed under the same conditions as described above except that 1500 ng/well of a biotin-F001 IgG (4th bleed) was used to detect the amount of bound antigen.

## RESULTS

U-937 Permeability Enhancing Activity.

The serum-free conditioned medium from U-937 cells in culture produced a positive response when tested in the Miles permeability assay. These results were unexpected since many cells do not produce a VPF-like activity as seen from Example 2, below. The Miles assay measures extravasation of Evan's blue dye-serum albumin complexes from the circulation after intradermal injection of a test material. However, the assay is non-specific and could measure positive response from a variety of substances, including, for example, histamine. It was therefore not known initially if the permeability enhancing activity produced by U-937 cells was related to guinea pig tumor VPF. To test this, the medium was mixed with an immunoadsorbent composed of protein A-Sepharose® and IgG obtained from polyclonal anti-sera against gVPF. Most, but not all, of the permeability enhancing activity present in the U-937 medium was adsorbed using this procedure, but not when control IgG obtained from rabbits not immunized with gVPF was used instead. Most of the permeability enhancing activity secreted into the medium of U-937 cells therefore appeared to be related to guinea pig tumor derived VPF. However, as discussed below, even though hVPF shares some immunocrossreactivity with gVPF, it is immunologically distinct from gVPF.

hVPF Purification.

Initial attempts at purification of U-937 cell derived permeability enhancing activity employed the purification method previously used for gVPF as described by Senger et al, Fed. Proc. 46, 2102 (1987). The application of this method, or minor modifications thereof, did not produce homogeneous protein from U-937 cell conditioned medium, even though the chromatographic behaviour of the permeability enhancing activity was very similar to that of gVPF. A novel purification method was therefore developed that incorporated cation exchange chromatography, metal affinity chromatography, and RP-HPLC (Fig. 1). In the first step, concentrated conditioned medium was passed over an S Sepharose cation exchange column (Fig. 2A). About 50-70% of the permeability enhancing activity was bound to the column at pH 7.0. The non-adsorbed activity was not characterized. The bound activity was eluted with a gradient of sodium chloride, and after concentration by ultrafiltration, loaded onto a metal affinity column (Fig. 2B). All detectable activity was tightly bound by the copper/IDA-Sepharose column, and was eluted after most of the other proteins in a gradient of imidazole. The final step utilized RP-HPLC (Fig. 2C) and resulted in elution of a group of  $M_r$  ~40 kDa proteins in the fractions associated with the peak of permeability enhancing activity (Fig. 2D). This method has been repeated numerous times and it reproducibly yielded  $M_r$  ~40 kDa protein of about 90% or greater purity as analyzed by SDS-PAGE with silver staining or by N-terminal sequence analysis. Approximately 1-2 µg of pure protein can be obtained per liter of U-937 cell conditioned medium.

Dose Response of hVPF Induced Permeability Enhancement.

Different amounts of hVPF were tested in the Miles permeability assay. The lowest dilution producing a positive response was at a hVPF concentration of about 2.75 nM. This corresponds to an injection dose of 22 ng, or 0.55 picomoles of  $M_r$  40 kDa hVPF. This is equivalent to only one-ninth the 200 ng required for a similar response by the gVPF described in U.S. Patent 4,458,550.

Amino Acid Sequence of hVPF.

hVPF was subjected to N-terminal amino acid sequence analysis (Table 1). Complete identity was observed between hVPF and the guinea pig tumor derived gVPF for the first 6 positions, but the sequence diverged for the next 4 amino acids sequenced. The identity is thus only 80% in this N-terminal region.

Table 1

Comparison of N-Terminal Sequences of hVPF and gVPF		
	Residue (pMole Yield)	
Cycle	hVPF	gVPF
1	Ala (827)	Ala (838)
2	Pro (427)	Pro (588)
3	Met (406)	Met (358)
4	Ala (130)	Ala (483)
5	Glu (85)	Glu (456)
6	Gly (55)	Gly (434)
7	Gly (78)	Glu (537)
8	Gly (150)	Gln (278)
9	Gln (34)	Lys (178)
10	Asn (29)	Pro (354)
Approximately 1 nmole (about 40 $\mu$ g as estimated by SDS-PAGE and silver staining) of hVPF was sequenced. The amino acids detected and the yield at each cycle (in parentheses) are shown. The average repetitive yield was 73%. These data are representative of several other runs on similar hVPF preparations.		

In order to obtain internal sequence information, hVPF was reduced, carboxymethylated with iodoacetic acid, and treated with trypsin. The resulting peptides were then separated using RP-HPLC. Several of the isolated peptides were sequenced, as shown in Table 2. None of these sequences, nor the N-terminal sequences, showed significant homology to proteins present in published data bases. The novel hVPF of the present invention thus is substantially different from previously described proteins and from gVPF.

Table 2

Sequence of hVPF Tryptic Peptides	
Peptide	
53	Phe Met Asp Val Tyr Gln Arg
71	Ile Phe Lys Pro Ser Cys Val Pro Leu Met Arg
93	Val Asp Ile Phe Glu Tyr Pro Asp
	Glu <sub>1</sub> Ile Glu Tyr <sub>8</sub>
Reduced and carboxymethylated hVPF was treated with trypsin and the peptides separated by RP-HPLC. Peptides that were well isolated were sequenced. The number designation corresponds to the fraction number in which the peptide appeared.	

Reactivity of hVPF With Anti-gVPF Antibodies.

Polyclonal rabbit antibody against gVPF was tested for cross-reactivity with hVPF using a sandwich ELISA. Figure 3 shows that hVPF is recognized by anti-gVPF IgG, but that the concentration of hVPF required for detection is about 10 fold higher than that for gVPF. Furthermore, although the maximum absorbance attained for hVPF (1.5 absorbance units above background) was significant, it was about 3 fold lower than that attained using gVPF as antigen. These results imply that the two proteins are related, but that hVPF is immunologically distinct from gVPF.

### Example 2

hVPF derived from U-937 cells was compared with VPF from several other cell lines by the Miles assay with the following results. The Mnnng HOS human osteogenic sarcoma cell line was used as representative of the prior art human cell lines for production of VPF described by Senger et al., *Cancer Res.* 48, 5629-5632 (1988). The cells were extracted at a density of  $1.5$  to  $2.0 \times 10^6$  cells/ml for 48 hours in serum-free basal medium, either RPMI 1640 or DME. The extract was collected, centrifuged to remove cells and debris, and then concentrated if necessary using Centricon 10 membranes (Amicon Corp.). The extracts were then tested in the Miles assay. The results as set forth in Table 3 demonstrate that the U-937 cells are outstandingly more effective as a producer of hVPF than the other human cell lines.

Table 3

Cell Line	Conditioned Media Conc'n.(a)	Miles Assay (units) (b)	Estimates of VPF Conc'n. (ng/ml) (c)
U-937(ATCC)			
Uncloned	1X	1	50
U-937(ATCC)	1X	2-8	100-400
Cloned (d)			
U-937(ATCC)			
Uncloned- Passaged through Nude mouse	1X	4-8	200-400
Mnnng HOS			
Human osteogenic sarcoma	10X	+ + + (e)	30 (e)
JURKAT			
Human acute lymphoblastic leukemia	12.3X	1-2	4-8
F2.11-2X			
Human T cell-T cell hybridoma	10X	0	<5
HFF			
Neonatal human foreskin fibroblasts	26.6X	0	<2
1MR 80			
Human fetal lung fibroblasts	26.6X	5	10
HEK			
Human embryonic kidney	16.9X	4	10
8387-M15			
Human sarcoma	36X	>16	>20

(a) Conditioned media were concentrated X-fold as indicated prior to testing by the Miles assay.

(b) Activity of sample = greatest dilution of sample at which blue spot was detectable.

1 unit (u) = the amount of VPF in a sample producing the smallest detectable blue spot discernible from control injections without VPF.

(c) Estimate of VPF concentration in the unconcentrated spent media in ng/ml based on 1 unit activity = 50 ng/ml, a relationship established with the guinea pig Line 10 VPF of U.S. Patent 4,458,550.

(d) Several clones, obtained by limiting dilution cloning procedures, produced activities in the 100-800 ng/ml range.

(e) This is an estimate based on the previously determined activity of Mnnng HOS cells, since dilutions of the samples were not performed in this assay.



Example 3Materials

All reagents and abbreviated designations, unless otherwise specified, are the same as described by Maniatis et al., Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1982. The  $\lambda$ gt10 library was obtained from Clontech Laboratories, Inc., Palo Alto, Calif. (Cat. No. HL 1036a), which was prepared from U-937 cells treated with phorbol myristate acetate (PMA).

MethodsOligonucleotide Screening of  $\lambda$ gt10 Clontech library.Methods.

Approximately  $5 \times 10^5$  clones from a  $\lambda$ gt10 cDNA library constructed from phorbol-ester stimulated U-937 cells (Clontech, Palo Alto, CA) were screened using two best-guess oligonucleotide probes. One probe, (5'-GTTCTGGCCGCCGCGCCCTCGGCCATAGGAGC-3'), which was based on N-terminal amino acid sequencing (amino acids 1-10 of hVPF) hybridized to a single gt10 clone. A second probe (5'-GTGGACATCTTCAGGAGTACCCCGACGAGATCGAGTAC-3') based on sequence information from a tryptic peptide corresponding to amino acids 33-45 of hVPF hybridized to seven clones containing inserts ranging in size from 0.8 to 3.5 kb. The screening procedure was that of Ullrich et al., EMBO J. 3, 361-364 (1984), with the following modifications: purified yeast tRNA (Sigma Cat. No. R3001) (0.01mg/ml) was used instead of salmon sperm and the sodium pyrophosphate and ATP were deleted from the hybridization solution. The largest of these clones, clone 4, which was 3.5 kb in size, was subcloned into pUC9 (New England Biolabs), thereby generating plasmid vector pMON3044 (see fig. 4) and 1195 bp of sequence was obtained using the general dideoxy-termination method of Sanger et al., Proc. Natl. Acad. Sci. USA 74, 5463-5467 (1977), employing reagents obtained from United States Biochemicals. The sequence corresponding to nucleotides 288 to 1110 was obtained by sequencing both strands of DNA. Nucleotides 147-288 were determined from only one strand, but were sequenced twice using two different sequencing primers.

DNA band migration patterns on agarose gels as a result of multiple restriction endonuclease digestion patterns of the pMON3044 construct indicated the orientation of the clone as shown in Fig. 4.

RESULTS

A 3.5 Kb cDNA clone (designated VPF-4) was isolated from a  $\lambda$ 10 library prepared from mRNA isolated from phorbol-ester stimulated U-937 cells. This clone was transferred to the plasmid vector pUC9 to thereby form construct pMON3044. Then 1195 bp of sequence for the hVPF cDNA was generated. A variety of restriction endonucleases were used to determine the orientation of the cDNA for hVPF in the plasmid vector. Subjecting the cDNA to the selected endonuclease treatment demonstrated that approximately 1300 and 1800 bp of untranslated nucleotide sequences for the amino and carboxy termini, respectively, exist in the hVPF cDNA. The size of the hVPF/pUC9 construct (see FIG. 4) is approximately 8200 bp. This is the sum of approximately 3500 bp from the hVPF cDNA and approximately 2700 bp from the plasmid pUC9.

The predicted cDNA sequence demonstrated 100% homology with the N-terminus and several tryptic peptides previously determined using direct amino acid sequencing. These sequences correspond to amino acids 1-18, 17-23, 33-45 and 48-55, respectively. A signal sequence (amino acids -26 to -1) and a single N-linked glycosylation site at Asn 75 were also found. These results confirm that hVPF is a secreted and glycosylated protein.

A further comparison between the amino acid sequences of hVPF and gVPF was made by comparing the N-terminal sequence of the U-937 cell-derived hVPF as shown above and the sequence of gVPF determined by amino acid sequencing of purified gVPF from guinea pig Line 10 tumor cells as follows:

5	hVPF	Ala	Pro	Met	Ala	Glu <sup>5</sup>	Gly	Gly	Gly	Gln <sup>1</sup>	Asn	His	His	Glu	Val	Val <sup>15</sup>
		*	*	*	*	*	*			*				*	*	*
10	gVPF	Ala	Pro	Met	Ala	Glu <sup>8</sup>	Gly	Glu	---	Gln <sup>9</sup>	Lys	Pro	Arg	Glu	Val	Val <sup>14</sup>
	hVPF	Lys	Phe	Met	Asp	Val <sup>20</sup>	Tyr	Gln	Arg	Ser	Tyr <sup>25</sup>	Cys	His	Pro	Ile	Glu <sup>30</sup>
		*	*	*	*	*	*		*	*	*	*		*	*	*
15	gVPF	Lys	Phe	Met	Asp	Val <sup>19</sup>	Tyr	Lys	Arg	Ser	Tyr <sup>24</sup>	Cys	Arg	Pro	Ile	Glu <sup>29</sup>
20	hVPF	Thr	Leu	Val	Asp	Ile <sup>35</sup>	Phe	Gln								
		*	*	*	*	*	*									
	gVPF	Met	Leu	Val	Asp	Ile <sup>34</sup>	Phe	Gln								

Comparison of the amino termini of VPF from guinea pig and human tumor cell lines demonstrate distinct differences. A gap between amino acid positions 7 and 8 in the gVPF sequence must be inserted for overlap to occur between the two proteins. Additionally, using numbering from hVPF, several areas of nonidentical residues are noted, namely at amino acids 7, 8, 10, 11, 12, 22, 27 and 31.

#### Example 4

##### Expression of Permeability-Enhancing Activity by C-127 Cells Transfected with the hVPF Gene.

Serum-free conditioned medium was collected from three selected hVPF cell lines (VPF-25B, -29B, and -30B) transfected with a BPV expression vector containing the hVPF cDNA sequence, or from a control line (BPV-1123) transfected with the BPV vector only. This BPV vector contains the entire bovine papilloma virus genome ligated to the well-known pBR322 derivative of pML2. Medium was concentrated 5-fold using a Centricon-10 ultrafiltration device before performing the Miles permeability assay. A positive response was indicated by the appearance of a blue spot at the site of intradermal injection of sample into the guinea pig, indicating extravasation of Evan's blue dye from the circulation.

The hVPF cDNA was thus expressed in mammalian cells with a bovine papilloma virus (BPV) vector. This vector is based on the 100% viral genome and utilizes the mouse metallothionein I promoter and the SV40 Late poly A addition site to regulate the expression of foreign genes. The vector was linearized at the unique BamHI site between the promoter and poly A addition site. The 5' overhanging ends of the vector fragment were filled in with Klenow enzyme and dNTP's. Similarly, the plasmid containing the hVPF cDNA was digested with XmaI and the 1021 bp VPF fragment was isolated by gel electrophoresis. This blunt end fragment was then inserted into the vector by ligation. Mouse C-127 cells were cotransfected with the hVPF expression vector and pSV2neo by the calcium phosphate precipitation method as described by Howley et al., *Meth. Enzymol.* 101, 387-403 (1983). G418 (genticin) resistant transfectants were selected. Colonies were picked and expanded into stable lines for assay by the Miles assay.

Various other examples will be apparent to the person skilled in the art after reading the present disclosure without departing from the spirit and scope of the invention. It is intended that all such examples be included within the scope of the appended claims.

## Claims

1. Human vascular permeability factor having an amino acid sequence substantially as follows:

Ala Pro Met Ala Glu Gly Gly Gly Gln Asn His His Glu Val  
 Val Lys Phe Met Asp Val Tyr Gln Arg Ser Tyr Cys His Pro  
 Ile Glu Thr Leu Val Asp Ile Phe Gln Glu Tyr Pro Asp Glu  
 Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu Met Arg  
 Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro  
 Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys  
 Pro His Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln  
 His Asn Lys Cys Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg  
 Gln Glu Lys Lys Ser Val Arg Gly Lys Gly Lys Gly Gln Lys  
 Arg Lys Arg Lys Lys Ser Arg Tyr Lys Ser Trp Ser Val Pro  
 Cys Gly Pro Cys Ser Glu Arg Arg Lys His Leu Phe Val Gln  
 Asp Pro Gln Thr Cys Lys Cys Ser Cys Lys Asn Thr Asp Ser  
 Arg Cys Lys Ala Arg Gln Leu Glu Leu Asn Glu Arg Thr Cys  
 Arg Cys Asp Lys Pro Arg Arg.

2. Human vascular permeability factor of Claim 1 which is N-glycosylated at amino acid residue Asn 75.  
 3. Human vascular permeability factor of Claim 1 which is non-glycosylated.  
 4. Human vascular permeability factor of Claim 1 including a signal sequence at the N-terminus as follows:

Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu  
 Ala Leu Leu Leu Tyr Leu His His Ala Lys Trp Ser Gln Ala.

5. The cDNA of human vascular permeability factor including a nucleotide sequence substantially as follows:

1 GCGCAGACAG TGCTCCAGCG CGCGCGCTCC CCAGCCCTGC CCGGCCCTCG  
 51 GCCGGGAGGA AGAGTAGCTC GCCGAGGCGC CGAGGAGAGC GGGCCGCCCC  
 101 ACAGCCCGAG CCGGAGAGGG ACGCGAGCCG CGCGCCCCCG TCGGGCCTCC  
 151 GAAACC ATG AAC TTT CTG CTG TCT TGG GTG CAT TGG AGC CTT  
 193 GCC TTG CTG CTC TAC CTC CAC CAT GCC AAG TGG TCC CAG GCT  
 235 GCA CCC ATG GCA GAA GGA GGA GGG CAG AAT CAT CAC GAA GTG  
 277 GTG AAG TTC ATG GAT GTC TAT CAG CGC AGC TAC TGC CAT CCA  
 319 ATC GAG ACC CTG GTG GAC ATC TTC CAG GAG TAC CCT GAT GAG  
 361 ATC GAG TAC ATC TTC AAG CCA TCC TGT GTG CCC CTG ATG CGA  
 403 TGC GGG GGC TGC TGC AAT GAC GAG GGC CTG GAG TGT GTG CCC  
 445 ACT GAG GAG TCC AAC ATC ACC ATG CAG ATT ATG CGG ATC AAA  
 487 CCT CAC CAA GGC CAG CAC ATA GGA GAG ATG AGC TTC CTA CAG  
 529 CAC AAC AAA TGT GAA TGC AGA CCA AAG AAA GAT AGA GCA AGA  
 571 CAA GAA AAA AAA TCA GTT CGA GGA AAG GGA AAG GGG CAA AAA  
 613 CGA AAG CGC AAG AAA TCC CGG TAT AAG TCC TGG AGC GTT CCC  
  
 655 TGT GGG CCT TGC TCA GAG CGG AGA AAG CAT TTG TTT GTA CAA  
 697 GAT CCG CAG ACG TGT AAA TGT TCC TGC AAA AAC ACA GAC TCG  
 739 CGT TGC AAG GCG AGG CAG CTT GAG TTA AAC GAA CGT ACT TGC  
 781 AGA TGT GAC AAG CCG AGG CGG TGA GCCGGGCAGG AGGAAGGAGC  
 825 CTCCTCAGG GTTTCGGGAA CCAGATCTCT CACCAGGAAA GACTGATACA  
 875 GAACGATCGA TACAGAAACC ACGCTGCCGC CACCACACCA TCACCATCGA  
 925 CAGAACAGTC CTTAATCCAG AAACCTGAAA TGAAGGAAGA GGAGACTCTG  
 975 CGCAGAGCAC TTTGGGTCCG GAGGGCGAGA CTCCGGCGGA AGCATTCCCC  
 1025 GCGGGGTGAC CCAGCACGGT CCCTCTTGGA ATTGGATTCT CCATTTTATT  
 1075 TTTCTTGCTG CTAAATCACC GAGCCCGGAA GATTAGAGAG TTTTATTCT  
 1125 GGGATTCTG TAGACACACC CACCCACATA CATACATTTA TATATATATA  
 1175 TATTATATAT ATATAAATTA A.

6. Plasmid hVPF/pUC9.

7. Human vascular permeability factor cDNA clone VPF-4 of about 3.5 kb including the cDNA of Claim

5.

8. Mammalian cells transfected with a vector containing the cDNA sequence for human vascular permeability factor.

9. C-127 mouse cells transfected with a vector containing the cDNA sequence for human vascular permeability factor.

10. Method for the production of human vascular permeability factor in vitro comprising growing cells derived from the human histiocytic lymphoma cell line U-837 in nutrient culture medium at about 35° to 38° C for a sufficient time to elaborate vascular permeability factor and isolating the resulting vascular permeability factor from the spent cells or the cell culture conditioned medium.

11. The method of Claim 10 in which the vascular permeability factor is isolated from the cell culture conditioned medium by

(a) cation exchange chromatography of said conditioned cell culture medium;

(b) metal affinity chromatography of the VPF active fractions from said cation exchange chromatography; and

(c) reverse phase HPLC of the active VPF fractions from said method affinity chromatography.

12. The method of Claim 10 in which the nutrient culture medium is serum-free.

13. The method of Claim 10 in which the cation exchange chromatography is carried out with a column of S-Sepharose Fast Flow cation exchange, the metal affinity chromatography is carried out with a copper/immunodiacetic acid (IDA)/Sepharose column, and the reverse phase HPLC is carried out with a C<sub>18</sub> reverse phase HPLC column.

14. A human vascular permeability factor having a molecular weight of about 34 to 42 kDa as determined by non-reduced SDS-PAGE and an N-terminal amino acid sequence as follows:

<sup>1</sup>Ala-Pro-Met-Ala-Glu-Gly-Gly-Gly-Gln-Asn.<sup>10</sup>

15. The human VPF of Claim 14 containing the following internal sequences:

(a) (Gln)Gln Gln Lys Pro,

(b) (Arg)Gln Glu Gln Arg(Pro Lys),

(c) Phe Met Asp Val Tyr Gln Arg(Arg),

(d) Ile Phe Lys Pro Ser Cys Val Pro Leu Met Arg, and

(e) Val Asp Ile Phe Gln Glu Tyr Pro Asp Glu Ile Glu Tyr.

16. The human VPF produced by the method of Claim 10.

## HUMAN VPF PURIFICATION SCHEME

U937 SERUM FREE CONDITIONED MEDIUM



CONCENTRATE 6X (AMICON)



S-SEPHAROSE FAST FLOW  
CATION EXCHANGE CHROMATOGRAPHY  
(NaCl GRADIENT ELUTION)



CONCENTRATE 20X (AMICON)



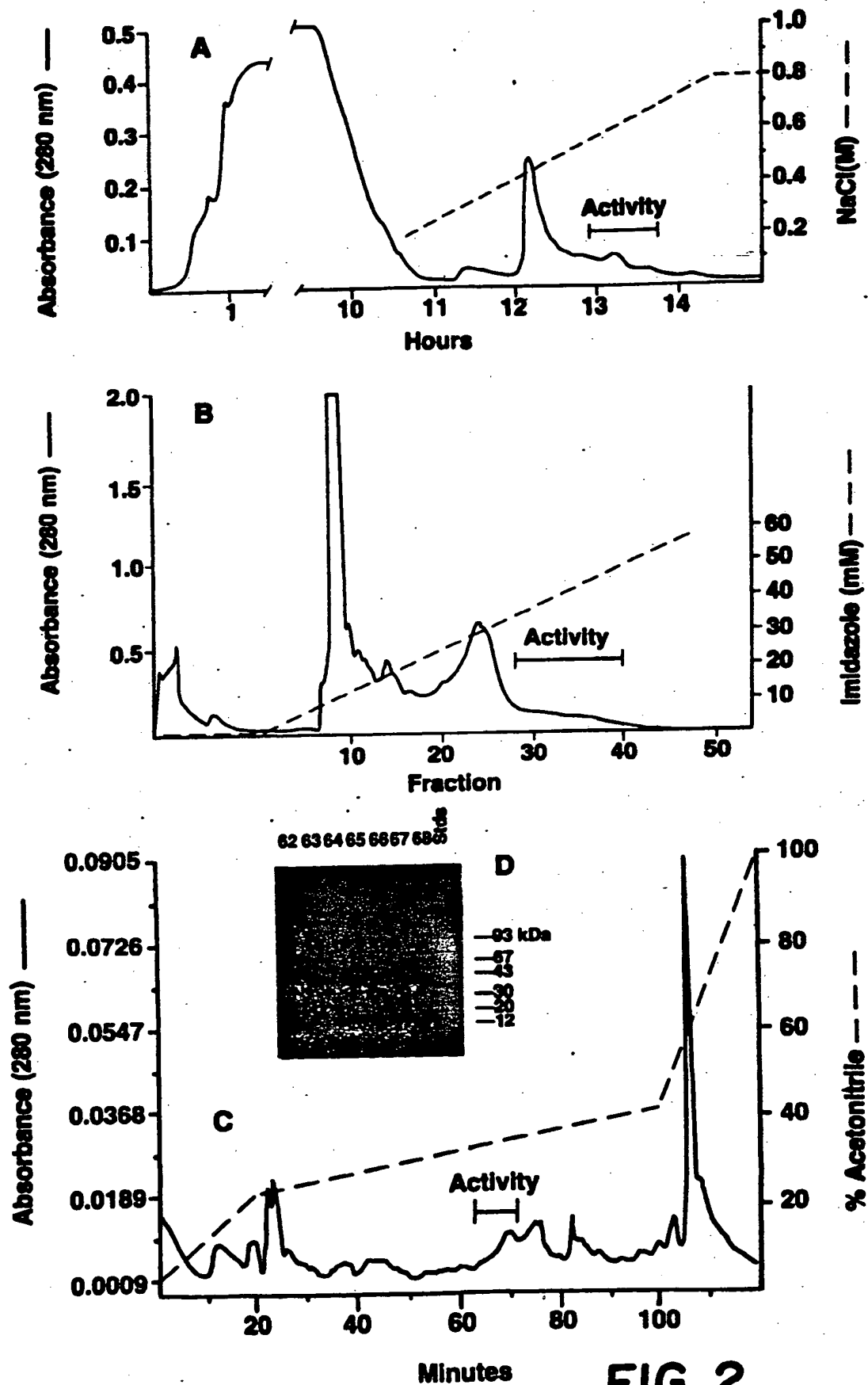
SEPHAROSE FAST FLOW  
IDA  $\text{Cu}^{++}$  CHELATE CHROMATOGRAPHY  
(IMIDAZOLE GRADIENT ELUTION)



$\text{C}_{18}$  RPHPLC  
( $\text{H}_2\text{O}$ /TFA/ $\text{CH}_3\text{CN}$  GRADIENT ELUTION)

YIELD:  $\sim 1 \mu\text{g/L}$  CONDITIONED MEDIUM  
PURIFICATION: 10,000-20,000 FOLD

**FIG.1.**



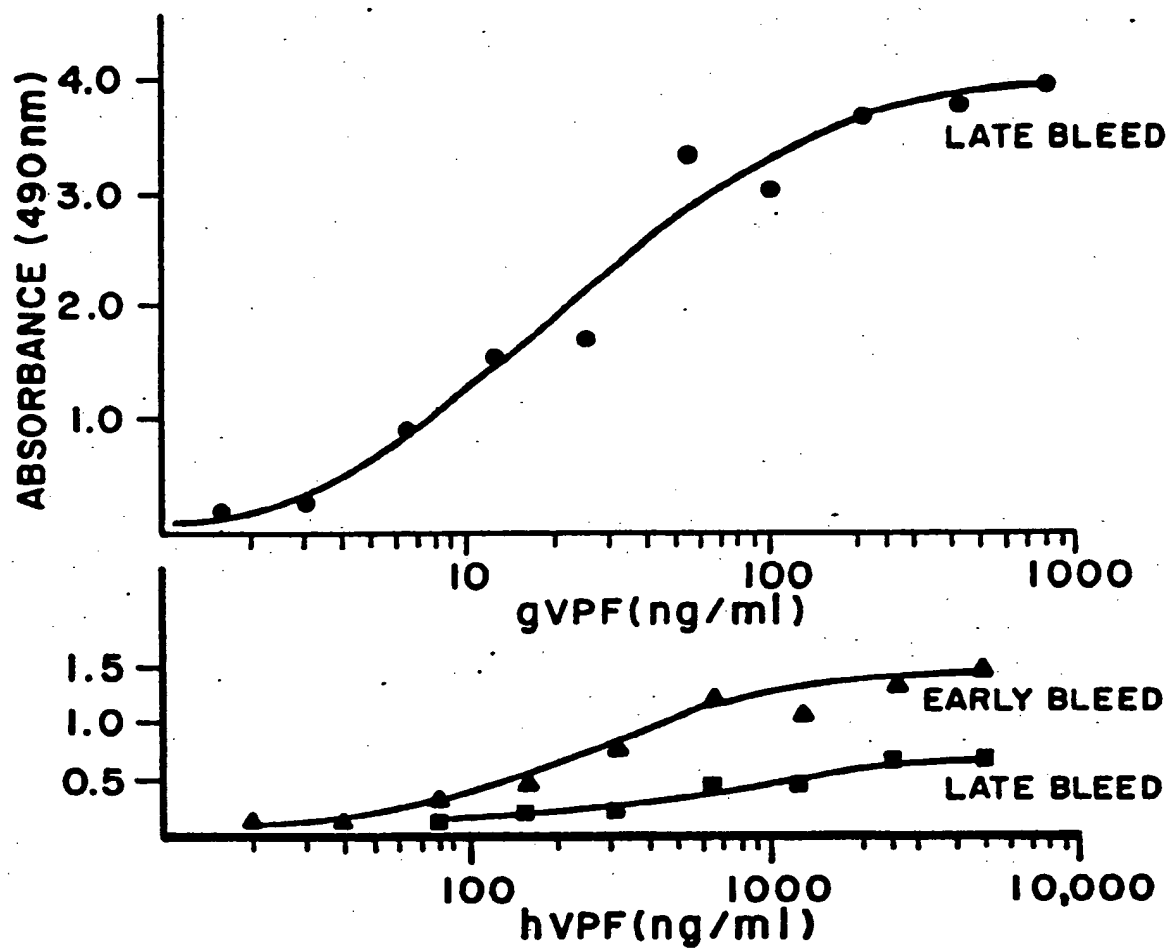


FIG. 3.



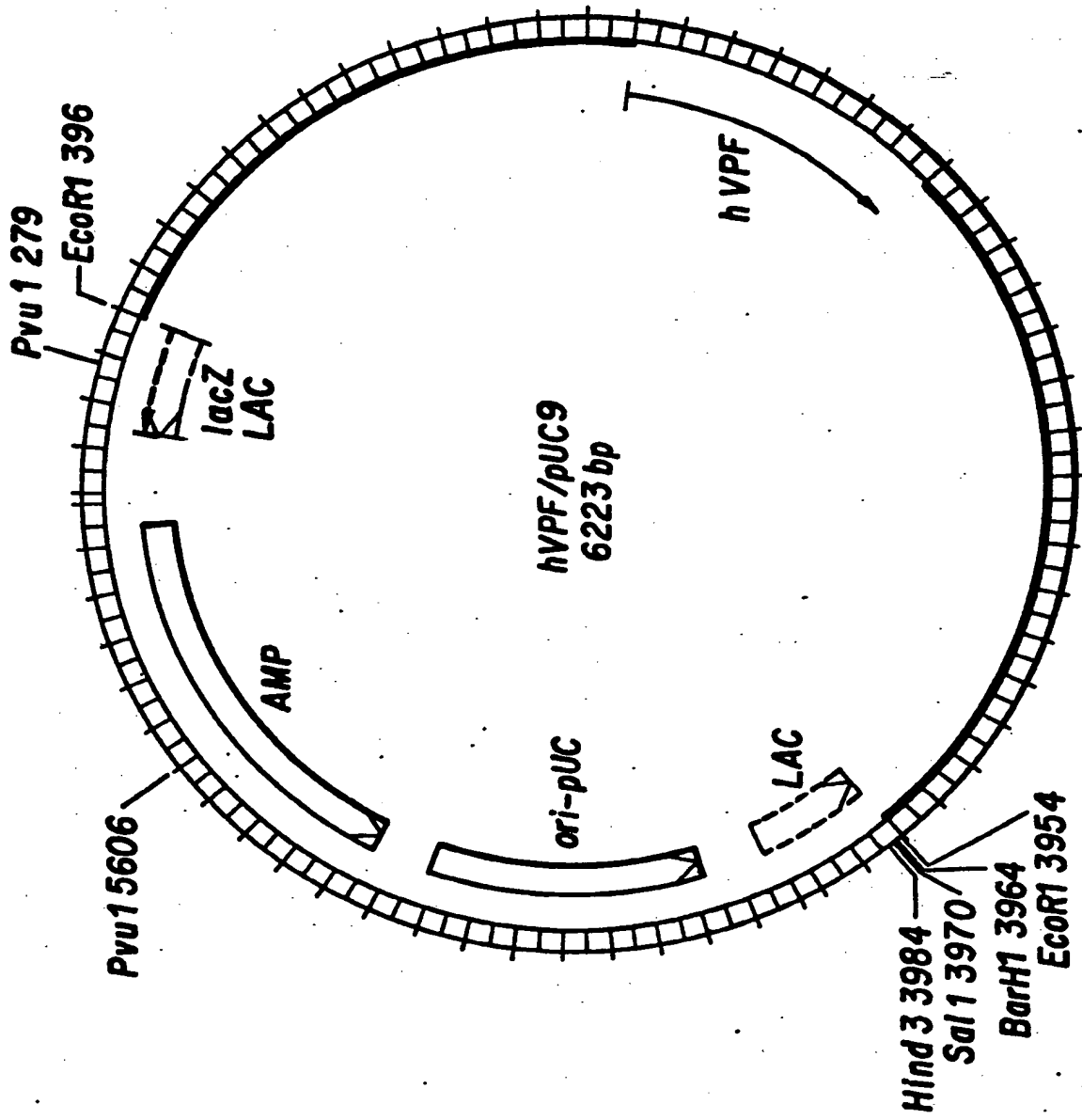


FIG. 4